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<b>(54) Title:</b> ARACHIDONIC ACID AND METHODS FOR THE PRODUCTION AND USE THEREOF  <b>(57) Abstract</b>  The present invention relates to processes for the production of arachidonic acid containing oils, which preferably are substantially free of eicosapentanoic acid. The invention also relates to compositions containing such oils, in an unmodified form, and to uses of such oils. In a preferred embodiment, <i>Pythium insidiosum</i> is cultivated, harvested and the oil is extracted, recovered, and used as an additive for infant formula.		

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ARACHIDONIC ACID AND METHODS FOR THE  
PRODUCTION AND USE THEREOF

5 This invention relates to the production of  
arachidonic acid, to compositions containing  
arachidonic acid and to uses thereof.

Arachidonic acid (ARA) is a long chain  
polyunsaturated fatty acid (PUFA) of the omega-6 class  
(5,8,11,14-eicosatetraenoic acid, i.e., 20:4). ARA is  
the most abundant C<sub>20</sub> PUFA in the human body. It is  
10 particularly prevalent in organ, muscle and blood  
tissues, serving a major role as a structural lipid  
associated predominantly with phospholipids in blood,  
liver, muscle and other major organ systems. In  
addition to its primary role as a structural lipid, ARA  
15 also is the direct precursor for a number of  
circulating eicosenoids such as prostaglandin E<sub>2</sub> (PGE<sub>2</sub>),  
prostacyclin I<sub>2</sub> (PGI<sub>2</sub>), thromboxane A<sub>2</sub> (TxA<sub>2</sub>), and  
leukotirenes B<sub>4</sub> (LTB<sub>4</sub>) and C<sub>4</sub> (LTC<sub>4</sub>). These eicosenoids  
exhibit regulatory effects on lipoprotein metabolism,  
20 blood rheology, vascular tone, leucocyte function and  
platelet activation.

Despite its importance to human metabolism, ARA  
cannot be synthesized in humans de novo. ARA is  
synthesized by the elongation and desaturation of  
25 linoleic acid (LOA), an essential fatty acid. This  
process requires the presence of the enzyme  $\Delta 6$ -  
desaturase, an enzyme present in the human body in low  
levels, Burre et al., Lipids, 25:354-356 (1990).

Accordingly, most ARA must be provided in the diet, and this is especially important during times of very rapid body growth, such as infancy.

During the first year of its life, an infant can  
5 double or triple its weight. Consequently, elevated levels of dietary ARA are required. To satisfy this increased demand, human breast milk contains high levels of ARA. Sanders et al., Am. J. Clin. Nutr., 31:805-813 (1978). ARA is the most prevalent C<sub>20</sub> PUFA  
10 in breast milk. Of those mothers, especially vegetarians, who do breast-feed their infants, many would benefit from additional dietary ARA. However, many mothers do not breast feed their infants, or do not breast feed for the entire period of rapid infant  
15 growth, choosing instead to utilize an infant formula.

No commercial infant formulas known to Applicant contain ARA. U.S. Patent No. 4,670,285 (Clandinin et al.), incorporated herein by reference, discloses the infant's requirement for fatty acids including ARA. To  
20 provide these fatty acids, Clandinin et al. suggest a blend of egg yolk, fish oil or red blood cell phospholipids and vegetable oils as the fat component of a proposed infant formula. However, fish oil contains high quantities of eicosapentaneic acid (EPA). EPA is known to depress ARA synthesis in  
25 infants. Carlson, et al., INFORM, 1:306 (1990). Thus, it would be desirable to be able to provide ARA without also providing additional EPA. Furthermore, egg yolks contain a relatively low concentration of ARA, such  
30 that Clandinin et al.'s mixture is not economically viable.

Because ARA is present in animal, but not vegetable, oils, its production in commercial quantities has remained a desirable, but elusive, goal.

Shinmen, et al., Microbiol. Biotech. 31:11-16 (1989), have reported the production of ARA by an isolated fungus, *Mortierella alpina*, using conventional stirred tank fermentation. (See also Japanese Patent 1,215,245 to Shinmen et al.). After culturing, the organisms are harvested, dried and their lipids extracted from the fungal biomass with an organic solvent and the lipids chemically (covalently) modified. For example, the lipid mixture is hydrolyzed or converted to ethyl esters and then combined with cyclodextrin prior to use as a dietary supplement. Shinmen et al. do not disclose or suggest the administration of unmodified microbial oils.

*Porphyridium cruentum*, a red microalgae, can be grown in ponds in large quantities and has a lipid content which can contain up to 40% ARA. Ahern, et al. Biotech. Bioeng. 25:1057-1070 (1983). Unfortunately, the ARA is primarily associated with galactolipids, a complex polar lipid not present in breast milk. Thus, not only is the total usable ARA produced a fraction of one percent of the biomass, but the form of the ARA is not suitable for use as an additive to infant formula without further modification.

U.S. Patent No. 4,870,011 (Suzuki et al.) discloses a method for obtaining lipids such as  $\gamma$ -linolenic acid from fungi of the genus *Mortierella*. The  $\gamma$ -linolenic acid is purified from the mixture of lipids contained in the fungi.

DE 3603000A1 (Milupa) discloses a highly polyunsaturated acid fat mixture and its use as the fat component of an infant formula. The fat mixture has a high content of ARA and docosahexanoic (DHA) acids in a ratio of 2.5:1 respectively, as well as a high content of cholesterol. Sources of the fatty acids are listed

as being certain types of macroalgae, fish oils, organ  
fats from beef and pork or highly refined egg yolk oil.  
A source of the DHA and ARA is said to be macroalgae of  
the phaeocophyte and rhodophyte types. There is no  
5 suggestion to use any microbes as a source of oil.  
Algal and fish oils also typically include EPA which  
depresses ARA synthesis in vivo. Additionally, highly  
refined egg yolk oil is not an economical source of  
ARA. Moreover, there is no disclosure therein of an  
10 ARA-concentrated additive for supplementing pre-  
existing infant formula.

Accordingly, there remains a need for an  
economical, commercially feasible method of producing  
ARA, preferably without concomitant production of EPA.  
15 It is an object of the present invention to satisfy  
that need.

It is a further object of the invention to provide  
an additive, and a source for that additive, for use in  
an infant formula such that the ARA levels in the  
20 formula approximate those levels in human breast milk.

It is an additional object of this invention to  
provide an ARA-containing fungal oil for use in  
enteral, parenteral or dermal products.

#### Summary of the Invention

25 This invention relates to the production and use  
of arachidonic acid containing fungal oil (ARASCO) and  
to compositions containing such oils. The oil can be  
referred to as a single cell oil. Fungi are cultivated  
under oil-producing conditions, harvested and the oil  
30 extracted and recovered. The oil, without further  
chemical modification, can be used directly to provide  
supplemental ARA to persons requiring such, including  
newborn infants, pregnant or nursing women or persons

exhibiting ARA-deficient pathologies. Advantages of the invention include its ease of production, and high purity, and lack of detectable amounts of EPA.

Detailed Description of the Preferred  
Embodiment of the Invention

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The present invention succeeds in providing an economical source of arachidonic acid (ARA). In one embodiment, this invention relates to a method for the production of an arachidonic acid-containing fungal oil (ARASCO) which is substantially free of eicosapentaneic acid (EPA). As used herein, "substantially free" means that the EPA is present in less than about one fifth of the amount of ARA in the oil. This oil, a single cell oil, can be administered directly, in an unmodified form. As used herein "unmodified" means that the chemical properties of the fatty acids, or the oils themselves, have not been covalently altered. Thus, for example, a temporary modification to the ARASCO or ARA which could be reversed following uptake of the oil would not be beyond the scope of this invention.

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Table 1. Fatty Acid Composition of Several Fungal Species.

Species	Fatty Acid									total fat
	14:0	16:0	16:1	18:1	18:2	18:3	20:4	20:5		
5										
<i>Mortierella alpina</i>	--	8.2	--	33.5	16.3	23.3	13.0	--	3.0	
<i>Mortierella elongata</i>	2.0	13.2	--	26.6	11.9	13.2	13.8	2.4	4.0	
<i>Mortierella isabellina</i>	0.3	15.7	0.8	55.8	11.1	9.0	--	--	7.3	
<i>Saprolegnia parasitica</i>	7.4	19.1	1.9	6.3	24.5	12.5	10.5	10.5	9.3	
<i>Pythium catenulatum</i>	6.5	9.9	10.3	21.2	18.5	3.5	13.4	10.9	5.0	
<i>Pythium coloratum</i>	13.6	9.9	--	14.7	10.9	2.5	24.3	21.7	2.2	
<i>Pythium gracile</i>	14.7	9.1	2.2	14.8	12.6	3.6	22.1	5.7	4.5	
<i>Pythium irregulare</i>	10.3	15.4	6.9	12.3	21.0	3.9	10.6	12.4	11.9	
<i>Pythium ultimum</i>	9.5	16.7	10.5	17.1	20.7	1.3	9.0	6.9	13.3	
<i>Pythium insidiosum</i>	9.5	11.4	12.1	1.0	8.3	9.3	31.9	--	2.8	
10										



Of those fungal species which previously have had their fatty acids characterized, it has been found that most do not make ARA. Weete, J.D., Fungal Lipid Biochemistry, Plenum Press, N.Y. (1974). Of those species which do make ARA, many, including all previously characterized *Pythium* species, produce significant quantities of eicosapentaenoic acid (EPA) in addition to ARA. Table 1 sets forth the fatty acid profile of *P. insidiosum* as well as the fatty acid profile of other species of fungi. Unexpectedly, it has been found that *P. insidiosum* produces ARA without concomitant production of EPA. As with fish oils, high EPA levels in dietary supplements result in a depression of the ability to form ARA from dietary linoleic acid (LOA). Accordingly, while those fungal species producing both ARA and EPA can be utilized in the process of this invention, it is preferable to use species which do not produce significant quantities of EPA. Such preferred species include *Pythium insidiosum* and *Mortierella alpina*. Both species are available commercially and are on deposit with the American Type Culture Collective in Rockville, Maryland, having accession numbers 28251 and 42430, respectively. Throughout this disclosure, unless otherwise expressly stated, *P. insidiosum* will be the representative fungal species.

One of the significant problems which an embodiment of the present invention overcomes, is the depression of ARA biosynthesis in infants caused by the presence of enhanced dietary levels of EPA. This problem can be corrected by providing ARA for use in infant formula at levels substantially similar to those found in human breast milk. Typically in human breast milk, the ratio of ARA:EPA is about 20:1 respectively.

The present invention specifically contemplates any microbial oil which provides a sufficient amount of ARA to overcome the negative effects of dietary EPA.

Preferably, the use of the ARA-containing oil will  
5 result in an ARA:EPA ratio of at least about 5:1. More preferably, the ratio will be at least about 10:1 and, most preferably, it will be at least about 20:1. As can be seen, the higher the amount of ARA in the end product, with respect to the amount of EPA, the more  
10 desirable is the result.

In a process of the present invention, the fungi are cultivated under suitable ARA-containing oil producing cultivating conditions. In general, techniques of fungal cultivation are well known to  
15 those of skill in the art and those techniques can be applied to the present inventive process. For example, cultivation of an inoculating amount of fungus can occur in submerged culture in shake flasks. The flask is provided with a growth medium, seeded with fungal  
20 mycelium, and grown on a reciprocating shaker for about three to four days.

The composition of the growth medium can vary but always contains carbon and nitrogen sources. A preferred carbon source is glucose, amounts of which  
25 can range from about 10-100 grams glucose per liter of growth medium. Typically about 15 grams/liter are utilized for shaker flask culture. The amount can be varied depending upon the desired density of the final culture. Other carbon sources which can be used  
30 include molasses, high fructose corn syrup, hydrolyzed starch or any other low cost conventional carbon source used in fermentation processes. Additionally, lactose can be provided as a carbon source for *P. insidiosum*. Thus, whey permeate, which is high in lactose and is a

very low cost carbon source, can be used as a substrate. Suitable amounts of these carbon sources can readily be determined by those of skill in the art. Usually, additional carbon needs to be added during the course of the cultivation. This is because the organisms use so much carbon that adding it all in a batch mode could prove unwieldy.

Nitrogen typically is provided in the form of yeast extract at a concentration of from about 2 to about 15 grams extract per liter of growth medium. Preferably, about four grams per liter are provided. Other nitrogen sources can be used, including peptone, tryptone, cornsteep liquor, etc. The amount to be added of these sources can easily be determined by those of skill in the art. Nitrogen can be added in a batch mode, i.e. all at one time prior to cultivation.

After cultivation for 3-4 days at a suitable temperature, typically about 25-30°C, an amount of fungi has grown which is sufficient for use as an inoculum in a conventional stirred tank fermentor (STF). Such fermentors are known to those of skill in the art and are commercially available. Fermentation can be carried out in batch, fed-batch, or continuous fermentation modes. Preferably, the STF is equipped with a Rushton-type turbine impeller.

The fermentor is prepared by adding the desired carbon and nitrogen sources. For example, a 1.5 liter fermentor can be prepared by mixing about 50 grams of glucose and about 15 grams of yeast extract per liter of tap water. As previously discussed, other carbon or nitrogen sources or mixtures thereof can be used.

The reactor containing the nutrient solution should be sterilized by, for example, heating prior to inoculation. After cooling to about 30°C, the inoculum

can be added, and cultivation initiated. Gas exchange is provided by air sparging. The air sparging rate can vary, but preferably is adjusted to from about 0.5 to about 4.0 VVM (volume of air per volume of fermentor per minute). Preferably the dissolved oxygen level is kept at from about 10% to about 50% of the air saturation value of the solution. Accordingly, adjustments in the sparge rate may be required during cultivation. Agitation is desirable. The agitation is provided by the impeller. Agitation tip speed preferably is set within the range of from about 50 cm/sec to about 500 cm/sec, preferably from about 100 to 200 cm/sec.

In general, the amount of inoculum can vary. Typically, from about 2% to about 10% by volume of inoculum can be used. Preferably, in a fermentor seed train about 5% by volume of inoculum can be used.

Nutrient levels should be monitored. When glucose levels drop below 5 g/l, additional glucose should be added. A typical cultivation cycle utilizes about 100 grams of glucose and about 15 grams of yeast extract per liter. It is desirable to deplete the nitrogen during the course of the cultivation as this enhances oil production by the fungi. This is especially true when *M. alpina* is used as the production organism.

Occasionally, the culture will produce an excessive quantity of foam. Optionally, an antifoaming agent, such as those known to those of skill in the art, e.g. Mazu 310®, can be added to prevent foam.

The temperature of cultivation can vary. However, those fungi which produce both ARA and EPA tend to produce less EPA and more ARA when cultivated at higher temperatures. For example, when *Mortierella alpina* is cultivated at less than 18°C, it begins to produce EPA.

Thus, it is preferable to maintain the temperature at a level which induces the preferential production of ARA. Suitable temperatures are typically from about 25°C to about 30°C.

5            Preferably, cultivation continues until a desired biomass density is achieved. A desirable biomass is about 25 g/l of the organism. Such a biomass typically is attained within 48-72 hours after inoculation. At this time, the organisms typically contain about 5-40%  
10        complex lipids, i.e. oil, of which about 10-40% is ARA, and can be harvested.

          Harvesting can be done by any suitable method such as, for example, filtration, centrifugation, or spray drying. Because of lower cost, filtration may be  
15        preferred.

          After harvesting, the mycelial cake can be extracted. The mycelial cake refers to the collection of biomass resulting after harvest. The cake can be loose or pressed, crumbled or uncrumbled. Optionally,  
20        the cake can have any residual water removed, as by vacuum drying or lyophilization, prior to extraction. If this option is selected, it is preferable to use nonpolar solvents to extract the ARA-containing oil. While any non-polar extract is suitable, hexane is  
25        preferred.

          Alternatively, the wet cake (which typically contains about 30-50% solids) can be crumbled and extracted directly using polar solvents such as ethanol or isopropyl alcohol, or supercritical fluid extraction  
30        with solvents such as CO<sub>2</sub> or NO. Preferably, the cakes are crumbled prior to extraction. Advantageously, the present invention permits the economical use of supercritical fluid extraction techniques. McHugh, et al., Supercritical Fluid Extraction, Butterworth

(1986). Such techniques are known to those of skill in the art and include those presently applied, for example, to decaffeinate coffee beans. While the yields from both wet and dry extractions are similar, wet extraction generally is a more economical process.

A preferable method of aqueous extraction involves mixing the mycelial biomass with the polar solvent isopropyl alcohol in a suitable reaction kettle. Such kettles are known. The use of three to six parts of solvent per part of biomass is desired. Most preferably, the mixing is done under nitrogen or in the presence of antioxidants to prevent the oxidation of the ARA in the lipid extract. As used herein "lipid extract", "oil", "lipid complex" and "fungal oil" are used interchangeably.

After extracting, the mixture can be filtered to remove the biomass from the solvent containing the lipid extract. At this point, the biomass can be recovered and used as a food supplement. As used herein, "food supplement" means feed or an additive to be mixed with typical feed, such as grain, etc., that can be provided to animals.

The solvent is separated from the lipid extract and also can be recovered for reuse, as by evaporation into a suitable collector, leaving what is referred to herein as the "crude oil." Use of isopropyl alcohol as the solvent desirably results in the removal of any residual water from the crude oil, as the evaporation removes the water/isopropyl alcohol azeotrope which has spontaneously formed.

While the crude oil can be used without further treatment, it also can be further purified. Processes such as those used in the preparation of lecithin from vegetable products, and known to those of skill in the

art, can be used in this additional purification step. Such processes do not chemically or covalently modify the ARA-containing lipids or the ARA itself.

Yields vary, but typically are about 5 grams of ARA-containing phospholipid per 100 grams of dried mycelia. In the case of *M. alpina*, an additional 10-30 grams of triglyceride per 100 grams of dry mycelia can be obtained. Either the crude oil or the refined product can be used for administration to humans. Both shall be included within the definition of ARASCO as used herein.

A most preferred object of the invention is to provide an additive for use with human infant formulas, such that the concentration of ARA in such formula closely approximates the concentration of ARA in human breast milk. Table 2 compares the composition of the fatty acids in ARASCO with those in breast milk and infant formula lacking and containing ARASCO.

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Table 2. Fatty acid composition of fungal oil products and mother's milk

<u>Fatty Acid</u>	<u>ARASCO</u>	<u>Infant formula<sup>1</sup></u>	<u>formula + oil</u>	<u>breast milk<sup>1</sup></u>
8:0	--	24.1	23.6	0.35
10:0	--	17.7	17.3	1.39
12:0	--	14.9	14.6	6.99
14:0	4.6	5.8	5.8	7.96
16:0	16.0	6.8	7.0	19.80
16:1	3.2	0.2	0.3	3.20
18:0	--	2.3	2.3	5.91
18:1	26.4	10.0	10.3	34.82
18:2n6	9.9	17.4	17.3	16.00
18:3n3	4.1	0.9	1.0	0.62
20:1	2.2	0.1	0.14	1.10
20:2n6	--	--	--	0.61
20:3n6	1.4	--	0.03	0.42
20:4n6	32.0	--	0.64	0.59
20:5n3	--	--	--	0.03
22:1	--	--	--	0.10
22:4n6	--	--	--	0.21
22:5n6	--	--	--	0.22
22:6n3	--	--	--	0.19

<sup>1</sup> Simopoulis, A., Omega-3 Fatty Acids in Health and Disease, pp.115-156 (1990)



As can be seen, the amount of ARA present in the infant formula supplemented by ARASCO closely approximates the ARA levels in human breast milk. Additionally, the total fatty acid composition of the infant formula has not been significantly altered by the addition of the ARASCO. Typically, between about 50 to about 1000 mg of ARASCO per liter of infant formula can be used. The specific amount of ARASCO required depends upon the ARA content. This can vary from about 10 to about 50% of the fatty acids in the oil. However, typically the ARA content is about 30%. When the ARA content is about 30%, an especially preferred supplementation rate is about 600 to 700 mg of ARASCO per liter of infant formula. Such a rate dilutes the pre-existing fat components of an infant formula such as Similac® (Ross Laboratories, Columbus, Ohio) by only one part ARASCO to fifty parts formula oils. Preferably, the ARASCO is substantially free of EPA.

When *Pythium insidiosum* is used in the described process, the extracted ARA-containing oil is predominantly phospholipid. When *Mortierella alpina* is used in this process, the ARA-containing oil is predominantly triglyceride. Both forms of ARASCO are useful as additives to infant formula. The former not only provides the formula with ARA, but also with an emulsifier, i.e., phosphatidyl choline, which is commonly added to commercial formulas. The oil from *M. alpina* is likely to be more economical to produce.

The ARA-containing oil of the present invention has many uses in addition to its use as an additive for infant formula. As known to those of skill in the art, there are many pathologies associated with ARA deficiencies, such as marasmus (Vajreswari, et al.,

Metabolism 39:779-782 (1990)) or atopic diseases (Melnik, B., Monatsschr. Kinderheilkunde, 138:162-166 (1990)). In one embodiment of the present invention, those pathologies are treated by administering a pharmaceutically effective amount of the oil of the present invention. The oil can be administered enterally, topically or parenterally, as selected by the provider of health care.

Encapsulation, as known by those of skill in the art, is an effective method of enteral administration. Capsules containing the fungal oil can be administered to those persons requiring or desiring dietary supplementation of ARA. Such a method is particularly effective for administering ARA to pregnant or nursing women.

In instances where ARASCO is being administered to combat ARA deficiency associated pathologies, a pharmaceutically effective amount should be administered. This amount can be determined by those of skill in the art without undue experimentation.

Another embodiment of the present invention entails cosmetic compositions containing ARASCO. Cosmetic compositions refer to those compounds applied as cosmetics. A preferred example of such a composition is a wrinkle cream. Such cosmetic compositions provide an effective means of topically applying ARA to skin to assist in maintaining skin tone.

The invention having been generally described, the following specific non-limiting examples are set forth to further illustrate the invention.

Example 1. Preparation of *P. insidiosum* lipid and addition to infant formula

In an 80 liter (gross volume) fermentor, 51 liters of tap water, 1.2 kg glucose, 240 grams of yeast extract and 15 ml of MAZU 210S® antifoam were combined. The fermentor was sterilized at 121°C for 45 minutes. An additional 5 liters of condensate water were added during the sterilization process. The pH was adjusted to 6.2, and approximately 1 liter of inoculum (at a cell density of 5-10g/l) of Pythium insidiosum (ATCC #28251) then was added. The agitation rate was adjusted to 125 RPM (250 cm/sec tip speed) and the aeration rate was set at 1 SCMF (standard cubic feet per minute). At hour 24 in the operation the aeration rate was increased to 3 SCFM. At hour 28 an additional 2 liters of 50% glucose syrup (1 kg glucose) were added. At hour 50 the fermentor was harvested, resulting in a yield of about 2.2 kg wet weight (approximately 15 g dry weight) per liter. Harvested biomass was squeezed to a high solids cake (50% solids) on a suction filter before freeze drying. The dried biomass was ground with a mortar and pestle and extracted with 1 liter of hexane per 200 grams of dry biomass at room temperature under continuous stirring for 2 hours. The mixture then was filtered and the filtrate evaporated to yield about 5-6 grams of crude oil per 100 grams of dry biomass. The biomass then was reextracted with 1 liter of ethanol per 20 grams of dry biomass for 1 hour at room temperature, filtered, and the solvent evaporated yielding an additional 22 grams of crude oil per 100 grams of dry biomass. The second fraction was predominantly phospholipids whereas the first fraction contained a mixture of phospholipids and triglycerides. The combined fractions produced an oil

containing about 30-35% arachidonic acid and no detectable EPA. This oil was added dropwise to the commercial infant formula product Simulac® (Ross Laboratories, Columbus, Ohio) at a supplementation rate of 600 mg per liter of prepared medium.

Example 2. Preparation of *M. alpina* lipid and addition to infant formula

*Mortierella alpina* (ATCC #42430) was grown in a 2 liter shake flask containing 1 liter of tap water and 20 grams of potato dextrose medium. The flask was under constant orbital agitation and was maintained at 25°C for seven days. After harvesting by centrifugation, the biomass was freeze dried yielding about 8 grams of lipid-rich mycelia. The mycelia was extracted using hexane as in example #1 and about 2.4g of crude oil resulted. This oil contains about 23% arachidonic acid and was added to the commercial formula Similac® dropwise in concentrations of 1000 mg per liter.

We claim:

1. A method for the production of an arachidonic acid-containing oil substantially free of eicosapentaneic acid, comprising:

- 5 (a) cultivating a species of *Pythium* which produces an arachidonic oil substantially free of eicosapentaneic acid under suitable oil-producing-cultivating conditions;
- (b) harvesting said *Pythium*;
- 10 (c) extracting said oil from said harvested *Pythium*, and
- (d) recovering said oil.

2. The method of claim 1, wherein said species comprises *Pythium insidiosum*.

3. Oil produced by a cultivated oil-producing species of *Pythium*, said oil containing arachidonic acid substantially free of eicosapentaneic acid.

4. The oil of claim 3, wherein said *Pythium* comprises *P. insidiosum*.

5. The method of claim 1, wherein said oil comprises at least about 10 parts arachidonic acid per part eicosapentaneic acid.

6. The method of claim 1, wherein said extraction comprises treating said harvested biomass with a supercritical solvent.

7. The method of claim 6, wherein said solvent comprises CO<sub>2</sub> or NO.

8. A method of providing arachidonic acid to an infant formula comprising adding an unmodified arachidonic acid-containing fungal oil to said infant formula.

9. The method of claim 8, wherein said fungus comprises a species of *Pythium* or *Mortierella*.

10. The method of claim 9, wherein said *Pythium* comprises *P. insidiosum* and said *Mortierella* comprises *M. alpina*.

11. The method of claim 8, wherein said fungal oil is substantially free of eicosapentanoic acid.

12. An additive for infant formula comprising an unmodified arachidonic acid-containing fungal oil.

13. The additive of claim 11, wherein said fungus comprises a species of *Pythium* or *Mortierella*.

14. The additive of claim 13, wherein said *Pythium* comprises *P. insidiosum* and said *Mortierella* comprises *M. alpina*.

15. A method of treating pathologies arising from arachidonic acid deficiencies in humans comprising administering a pharmaceutically effective amount of an unmodified arachidonic acid-containing fungal oil to a human in need of such treatment.

16. The method of claim 15, wherein said fungal oil is extracted from a species of *Pythium* or *Mortierella*.

17. The method of claim 16, wherein said *Pythium* comprises *P. insidiosum* and said *Mortierella* comprises *M. alpina*.

18. The method of claim 17, wherein said fungal oil is administered enterally.

19. The method of claim 18, wherein said oil is substantially free of eicosapentaenoic acid.

20. The method of claim 17, wherein said fungal oil is administered topically.

21. The method of claim 17, wherein said fungal oil is administered parenterally.

22. The method of claim 18, wherein said method of administration comprises administering an

encapsulated form of arachidonic acid containing fungal oil.

23. The method of claim 18, wherein said human comprises a pregnant or nursing woman.

24. The method of claim 22, wherein said human comprises a pregnant or nursing woman.

25. Cosmetic compositions comprising arachidonic acid-containing fungal oils.

26. The composition of claim 25, wherein said oil is from a fungus comprising a species of *Pythium* or *Mortierella*.

27. The composition of claim 25, wherein said *Pythium* comprises *P. insidiosum* and said *Mortierella* comprises *M. alpina*.

28. The composition of claim 27, wherein said composition comprises a wrinkle cream.

29. A food supplement comprising the extracted *Pythium* of claim 1.

30. A method of producing an arachidonic acid-containing fungal oil from the species *Pythium insidiosum*, comprising:

- 5 (a) cultivating *P. insidiosum* in a stirred tank fermentor containing glucose and yeast extract to achieve a desired biomass containing said fungal oil;
- (b) harvesting said biomass,
- (c) extracting said fungal oil from said biomass by mixing said biomass with a polar solvent, such that  
10 said solvent extracts said oil from said biomass,
- (d) separating said solvent containing said extracted oil from said biomass, and
- (e) recovering said oil from said solvent.

31. The process of claim 30 further comprising further purifying said oil.

32. The process of claim 31, wherein said fermentor has a sparge rate of from about 0.5 VVM to about 2.0 VVM and an agitation tip speed of from about 50 cm/sec to about 350 cm/sec.

33. The process of claim 30, wherein said harvesting is done by filtration.

34. The process of claim 33, wherein said solvent comprises isopropyl alcohol.

35. The process of claim 34, wherein about 3 parts solvent per part biomass are added.

36. The process of claim 35, wherein said oil is recovered by evaporation of said solvent.

37. The oil produced by the process of claim 30.

38. An additive for infant formula comprising the crude oil of claim 37.

39. The additive of claim 38, wherein said oil comprises at least about five times more arachidonic acid than eicosapentanoic acid.

40. A method for treating pathologies arising from arachidonic acid deficiencies comprising administering a pharmaceutically effective amount of the oil of claim 37.

41. Cosmetic compositions comprising the oil of claim 37.

42. The cosmetic composition of claim 41, comprising a wrinkle cream.

43. A process for the production of arachidonic acid-containing oil substantially free of eicosapentanoic acid from *Pythium insidiosum* comprising;

- 5 (a) cultivating said *Pythium* in a fermentor in the presence of growth sustaining amounts of glucose and yeast extract until a desired amount of biomass is produced,



- 10           (b) harvesting said biomass,  
             (c) drying said biomass,  
             (d) extracting said dried biomass with a non-  
polar solvent to produce a solvent/oil mixture,  
             (e) separating said biomass from said solvent/oil  
mixture, and  
15           (f) recovering said oil from said solvent.

44. The process of claim 43 further comprising reextracting said biomass with a polar solvent, and repeating steps (d) through (f) to recover additional oil from said polar solvent.

45. The process of claim 43, wherein said fermentor has a sparge rate of from about 0.5 to about 2.0 VVM and an agitation tip speed of from about 50 cm/sec to about 350 cm/sec.

46. The process of claim 43, wherein said harvesting is done by filtration.

47. The process of claim 44, wherein said non-polar solvent comprises hexane.

48. The process of claim 47, wherein about 3 parts solvent per part biomass are added.

49. The process of claim 48, wherein said oil is recovered by evaporation of said solvent.

50. The process of claim 47, wherein said polar solvent comprises ethanol.

51. The oil produced by the process of claim 43.

52. An additive for infant formula comprising the crude oil of claim 51.

53. The additive of claim 52, wherein said oil comprises at least about 5 parts arachidonic acid per part eicosapentanoic acid.

54. A method for treating pathologies arising from arachidonic acid deficiencies comprising

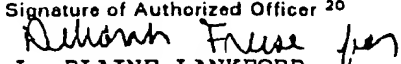
administering a pharmaceutically effective amount of the oil of claim 51.

55. Cosmetic compositions comprising the oil of claim 51.

56. The cosmetic composition of claim 55, comprising a wrinkle cream.

# INTERNATIONAL SEARCH REPORT

International Application No. PCT/US92/00517

<b>I. CLASSIFICATION OF SUBJECT MATTER</b> (if several classification symbols apply, indicate all) <sup>3</sup>		
According to International Patent Classification (IPC) or to both National Classification and IPC		
IPC (5) : C12P 7/64; A23C 9/00; A61K 6/00, 7/00 US CL : 435/134; 424/401; 426/585		
<b>II. FIELDS SEARCHED</b>		
Minimum Documentation Searched <sup>4</sup>		
Classification System	Classification Symbols	
U.S.	435/134, 424/401, 426/585	
Documentation Searched other than Minimum Documentation to the extent that such Documents are included in the Fields Searched <sup>5</sup>		
<b>III. DOCUMENTS CONSIDERED TO BE RELEVANT</b> <sup>14</sup>		
Category*	Citation of Document, <sup>16</sup> with indication, where appropriate, of the relevant passages <sup>17</sup>	Relevant to Claim No. <sup>18</sup>
Y	JP, A, 1,196,255 (SUNTORY, LTD) 08 AUGUST 1989. SEE WPI ABSTRACT NO. 89-268287/37	1-56
Y	JP, A, 1,215,245, (SUNTORY, LTD) 29 AUGUST 1989. SEE WPI ABSTRACT NO. 89-290735/40.	1-56
Y, E	US, A, 5,089,269, (NODA ET AL) 18 FEBRUARY 1992. SEE EXAMPLE 3-6.	25-28, 55-56
A, P	US, A, 5,013,569, (RUBIN) 07 MAY 1991. SEE ENTIRE DOCUMENT.	1-56
A	US, A, 4,526,793, (INGENBLEEK ET AL) 02 JULY 1985. SEE ENTIRE DOCUMENT.	1-56
<p>* Special categories of cited documents:<sup>15</sup></p> <div style="display: flex; justify-content: space-between;"> <div style="width: 48%;"> <p>"A" document defining the general state of the art which is not considered to be of particular relevance</p> <p>"E" earlier document but published on or after the international filing date</p> <p>"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)</p> <p>"O" document referring to an oral disclosure, use, exhibition or other means</p> <p>"P" document published prior to the international filing date but later than the priority date claimed</p> </div> <div style="width: 48%;"> <p>"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention</p> <p>"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step</p> <p>"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art</p> <p>"&amp;" document member of the same patent family</p> </div> </div>		
<b>IV. CERTIFICATION</b>		
Date of the Actual Completion of the International Search <sup>2</sup>		Date of Mailing of this International Search Report <sup>2</sup>
13 MARCH 1992		05 MAY 1992
International Searching Authority <sup>1</sup>		Signature of Authorized Officer <sup>20</sup>
ISA/US		 L. BLAINE LANKFORD

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